Result verification, code verification and computation of support values in phylogenetics

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Abstract
Verification in phylogenetics represents an extremely difficult subject. Phylogenetic analysis deals with the reconstruction of evolutionary histories of species, and as long as mankind is not able to travel in time, it will not be possible to verify deep evolutionary histories reconstructed with modern computational methods. Here, we focus on two more tangible issues that are related to verification in phylogenetics (i) the inference of support values on trees that provide some notion about the 'correctness' of the tree within narrow limits and, more importantly; (ii) issues pertaining to program verification, especially with respect to codes that rely heavily on floating-point arithmetics. Program verification represents a largely underestimated problem in computational science that can have fatal effects on scientific conclusions.

Keywords: phylogenetics; support values; program verification; maximum likelihood; bootstrap; software bug

INTRODUCTION
The goal of the discipline that emerged in the late 1960s [1, 2] and is now called phyloinformatics consists of reconstructing phylogenetic (evolutionary) trees from morphological or molecular sequence data. The phylogenetic history of a set of \( n \) organisms (represented, for instance, by their DNA-sequence data) is typically depicted as a fully bifurcating (strictly binary) tree topology. A tree-like model of evolution may be an over-simplification of the evolutionary processes, in particular in the presence of lateral gene transfer [3, 4], but in contrast to networks, trees are more straight-forward to visualize, understand, interpret and handle in a mathematical and computational framework. The \( n \) extant (currently living) organisms of the input data set [usually a multiple sequence alignment (MSA)] are located at the leaves (tips) of such a tree, while the inner nodes represent extinct common ancestors. If one is willing to accept the hypothesis of a tree-like evolutionary process, two key questions arise: what is the order of magnitude for the number of potential tree topologies with \( n \) taxa, and based on which criteria should we chose the ‘best’ among all potential tree topologies, that is, the tree that best fits our input data. In fact, the number of potential tree topologies increases super-exponentially with the number of organisms \( n \) according to: \( \prod (2i - 5) \) for \( i = 3, \ldots, n \) (this can be easily derived from the analogous formula for rooted binary trees proposed in [5]). As a consequence, for most common tree-scoring criteria such as parsimony [1, 6] or likelihood [7], finding the best tree is a NP-hard optimization problem [8, 9] because they require an exhaustive search of tree space. Thus, heuristics need to be deployed for finding the

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best-known parsimony or likelihood tree. At present, no mathematical tools are available to determine the score of the best possible tree (given a data set), as measured by parsimony or likelihood criteria. Moreover, for trees inferred with heuristic search strategies under maximum likelihood, there does not exist a quality guarantee, for instance, that the score of the tree returned is at most 10% worse than the score of the best tree. As such one can only compare the relative performance of different search strategies on real biological data sets. By using benchmark data sets, one can empirically observe that one search strategy yields better scores than another.

Alternatively, one can assess the ability of the implementations of different models and heuristic tree reconstruction strategies such as parsimony [10], maximum likelihood [11–13] or Bayesian approaches [14, 15] to recover the ‘true’ tree on simulated data sets. This approach is best used to assess questions with respect to input data set assembly and shape [16–22] (see ref. [23] for a summary of accuracy assessment techniques in phylogenetics). For this type of verification, one assumes a given true tree topology generated either with real biological data or by a computer program [24]. Then, a MSA is generated via a Markov process by letting an ancestral sequence evolve along the branches of the tree according to some statistical model of sequence evolution [25–27]. This will produce a MSA that can be used as input for phylogenetic tree reconstruction methods. The topological distances (see ref. [28] for a tool implementing most common tree distance metrics) between the trees that were reconstructed by the inference methods and the true tree that generated the data can then be deployed to assess the ability of the methods under consideration to reconstruct the true tree. This approach however is not without caveats. For example, there is no guarantee that the implementation of the simulated alignment generation programs is correct (e.g. consider the well-documented bug history of the widely used simulated data generation program Seq-Gen [25] at http://tree.bio.ed.ac.uk/software/seqgen/). Furthermore, the sequences along the true tree are typically generated using one of the common models of statistical sequence evolution, such as, for instance, the widely used [29] General Time Reversible (GTR [30]) model of nucleotide substitution with the $\Gamma$ model of rate heterogeneity [31]. In turn, the model that was used to generate the data will then also be used to reconstruct the tree. Thus, the true model of evolution is known a priori, and all the potentially simplistic assumptions, for instance, that the model is time-reversible and that sites (alignment columns) evolve independently, are already incorporated into the simulated data.

In other words, phylogenetic tree inference on simulated data (assuming a perfect model) tends to be easier than on real data. For instance, the assessment of the original PHYML [20] code for likelihood-based phylogenetic inference was mainly based on simulated data, and showed that PHYML performed as well as other programs. An assessment using real data then revealed that other heuristic search strategies that explore the tree search space more thoroughly returned trees with significantly better (in the statistical sense) likelihood scores [32]. Thus, because real biological data is less perfect, optimization tends to be harder on real biological data than on simulated data that has been generated using a known model. Nonetheless, simulated data can be used to test the behaviour of search algorithms and their ability to recover the ‘true’ tree in a best case scenario, that is, when the model perfectly fits the data.

Until recently, the incorporation of insertion and deletion events into simulated alignments represented a problem. However, new tools such as DAWG [27] and Indelible [26] now provide for simulated data generation under a more realistic insertion/deletion model. Another potential problem that can occur on data sets with a relatively small number of alignment sites, is that the true tree is not necessarily the maximum likelihood tree [22]. This is a result of maximum likelihood being consistent in the statistical sense, that is, the reconstructed tree converging to the true tree when the number of sites goes to infinity [33]. By using simulated data experiments (where model misspecification is not an issue) and RAxML for tree reconstruction, we show that, especially on short alignments, RAxML consistently finds trees that have a better log-likelihood score than the true tree. This does not necessarily mean that the inferred ML estimates on the short alignments are incorrect or of bad quality with respect to their topological distance to the true tree. However, as shown in Figure 1, the difference between the log-likelihood scores of the true tree and the inferred trees as well as the topological distances between the true tree and
the estimated trees decrease with alignment length (alignment width).

We generated simulated alignments using DAWG containing between 768 and 12,799 sites on a real biological tree with 1908 organisms. We optimized ML model parameters (branch lengths, GTR and \( \Gamma \) model parameters) using RAxML (\(-f e\) option) on the true tree and the inferred trees to compute the ratios between the log-likelihood scores. RAxML was also used to compute the symmetric topological difference (frequently also denoted as RF distance, albeit this terminology is not entirely correct; [33]) between the inferred trees and the true tree.

Based on the prolegomena, it is generally impossible to verify tree reconstruction programs with respect to their ability to infer the true or a reasonably correct tree because for real biological data the true tree and evolutionary model (if evolution is tree like) is commonly not known and computational verification experiments based on simulated data may represent easy tree reconstruction problems because the simulated data has been generated according to our simplistic models. Also no guarantee exists that the implementation of simulated data generation programs is correct.

Finally, the method deployed for computing a MSA can also have a significant impact on the tree reconstruction process [34] and ideally one would like to simultaneously infer trees and MSAs [35–37] which represents a hard computational problem and also a difficult modelling problem, because, at present, it is not clear which criterion to use for assessing MSA optimality. In phylogenomics, issues pertaining to a lack of a broadly accepted criterion for orthology assignment (see [38–40] for alternative approaches), gappy data [41] (sequence data for every organism and every gene under study is typically not available) and the discordance between gene trees and the species tree [42, 43] lead to the introduction of additional errors at the stages of input data assembly and alignment as well as the actual phylogenetic inference step.

Thus, given all of the aforementioned problems, verifying phylogenetic inference approaches is practically impossible. However, phylogenetic inference methods may be verified empirically, that is, if they can be successfully deployed to improve our living conditions, by using phylogenetic information to develop new drugs [44] or to disentangle [45] and predict viral outbreaks. Therefore, in the following we will briefly address two more tangible verification problems: the inference of support values on trees and the verification of codes for phylogenetic inference.

**Inference of Support Values**

The inference of support values on un-rooted binary-tree topologies intends to solve the following problem: Given a tree with five organisms A, B, C, D, E as depicted in Figure 2 (one may assume that
this is the best-known maximum likelihood tree), we want to determine a degree of confidence for the organisms being split into the subgroups that are induced by cutting the tree at an inner branch. In fact, we want to compute such a confidence value for every inner (internal) branch of the tree. Such splits of the taxon set into two disjoint taxon sets that are induced by inner branches of the tree are also termed non-trivial bipartitions. A trivial bipartition is a cut/split of the tree at a branch leading to a terminal organism, for instance, at the branch leading to organism A in Figure 2. Those bipartitions are called trivial because they do not provide any information about the tree topology, that is, the bipartition A|BCDE is contained in all possible 15 unrooted tree topologies for five organisms. Note that, the set of all \( n - 3 \) non-trivial bipartitions of an un-rooted binary tree (AB|CDE and ABC|DE in the example) with \( n \) taxa suffices to fully characterize the tree topology. In other words, the list of non-trivial bipartitions and the tree topology are equivalent representations of the same mathematical object. A common misconception among biologists is that support values are assigned to nodes of the tree, rather than to inner branches.

A list of non-trivial bipartitions as induced by a set of trees (e.g. a collection of plausible trees that have been sampled using Bayesian methods [14, 15, 46] or a collection of bootstrap replicate trees; see below) and their respective frequency of occurrence therein can then, for instance, be used to assign confidence values to the inner branches of a best-known ML tree or to reconstruct a consensus tree. It is important to be aware that, given all of the aforementioned limitations regarding verification in phylogenetics, those bipartition support values do not provide an indication of whether a true evolutionary split of the taxa has been recovered. Nonetheless, bipartition support values are commonly interpreted as a measure for the correctness of bipartitions.

To date, the most widely used techniques are: Bootstrapping [47], likelihood ratio-tests [13, 48] (a modification of the standard likelihood-ratio test [49]), and posterior probabilities as obtained by MCMC-based Bayesian methods [50] (see [51, 52] for reviews on support values).

**Bootstrapping**

The general bootstrap procedure was introduced by Brad Efron in 1979 [53] as a means to infer the variability of an unknown distribution for an estimator \( T \) using computer-based methods. The phylogenetic bootstrap procedure (Figure 3) was proposed in 1985 by Joe Felsenstein [47]. The underlying idea of the phylogenetic bootstrap is to assemble a certain number \( r \) of so-called bootstrap replicate alignments by randomly drawing sites (with replacement) from the original MSA. So, each bootstrapped alignment will contain exactly the same number of sites as the original alignment, but exhibit a slightly different site composition. Then, a tree is reconstructed for each of the \( r \) replicates using the estimator \( T \) (the tree reconstruction algorithm of choice), such that one obtains a set of \( r \) (potentially) distinct tree topologies. Those trees, that can be represented as a list of bipartitions, can then be used to build a strict or majority rule consensus tree as originally proposed in [47]. From the \( r \) replicates, one builds a in most cases multifurcating tree containing all bipartitions that occur in all \( r \) (strict) or more than \( r/2 \) (majority rule) trees in the set of replicates (see Figure 4, for an example).

Alternatively, one can draw bipartition support values on the best-known ML tree obtained from...
the original alignment. To do this, one counts how frequently a bipartition of the best-known ML tree occurs in the r bootstrap trees and assigns the frequency of occurrence to the bipartition.

A persistent issue is that the tree inference problem on each bootstrap replicate is also NP-hard. Hence, unlike in general applications of the bootstrap, our estimator T (whose quality is unknown) is only an approximation of the exact estimator \( T_0 \) (an algorithm that finds ‘the’ optimal ML tree).

One major disadvantage of the bootstrap is that, it is computationally expensive. Instead of conducting a small number of tree inferences, we need to compute between 100 and 1000 replicates. In fact, it is not clear how many bootstrap replicates \( r \) may be required to obtain reliable support values, albeit some theoretical [54] and practical [55] suggestions exist. The number of required replicates appears to be closely related to the behaviour of the estimator \( T \) on the specific data set being analyzed [55]. If the phylogenetic signal in the data is strong, relatively few replicates (\( r = 100 \)) are required and the bootstrap procedure will return a large number of identical trees. If the signal is weak (pointing towards multiple local optima in the likelihood landscape), a large number of replicates (\( r = 1000 \)) is required [55] (each replicate returning a different tree) to obtain stable majority rule trees (i.e. consensus trees that do not change if more replicates are computed). At present this is only an empirical observation that should be further investigated.

**Likelihood-ratio tests**

Approximate likelihood-ratio tests have been proposed [48] as a fast alternative to bootstrapping and implemented in programs such as PHYML v3.0 [13], FastTree v2.1 [56] and RAxML v7.2.8 [11]. In principle, they work as follows: Given a maximum likelihood tree that is locally optimal with respect to the application of NNI (nearest neighbour interchange) moves, the algorithm visits one inner branch at a time and computes three log likelihood values \( L_0, L_1 \) and \( L_2 \). \( L_0 \) is the log likelihood of the NNI-optimal tree, and \( L_1 \) and \( L_2 \) are the log likelihoods of the two alternative trees obtained by applying the two possible NNI interchanges to the branch under consideration (Figure 5). Then, \( L_0 \) and \( L_{\text{alt}} \) (best of the two alternative values \( L_1 \) and \( L_2 \)) are used to compute support statistics (see [48] for details). Instead of simply using \( L_0 \) and \( L_{\text{alt}} \) as originally proposed in [48], current implementations [11, 13, 56] use a Shimodaira Hasegawa-like test [13, 57], that essentially relies on random re-sampling of the per-site log-likelihood scores by using between 100 and 1000 samples. While this method is still very fast and seems to yield support values that are mostly in agreement with other bipartition support measures [13, 56], the interpretation of bootstrap support values remains difficult (see the ‘Introduction’ in [48] that entails additional references, [58, 59] and pp. 346–354 in [33]). Any comparison of aLRT versus bootstrap support values may be debatable, and there is one key issue that may require further investigation. The NNI move that is executed to obtain the three
likelihood values $L_0$, $L_1$ and $L_2$ (or SH-like re-sampled values for that matter) is only a local move and does not guarantee that the two best log-likelihood scores used in the test for that inner branch have been found. This is especially problematic for large trees with thousands of taxa. Thus, in comparison to the standard phylogenetic bootstrap or Bayesian posterior probabilities (see below), the immensely large tree space is explored to a significantly smaller degree for obtaining support values. This may potentially influence the quality of the support values obtained by these tests. It would therefore be desirable to assess the impact of more rigorous topological moves such as SPR (sub-tree pruning re-grafting) or TBR (tree bisection reconnection) moves on approximate LRT-support values. Therefore, it may be good practice to use at least one method (posterior probabilities or bootstrapping) that deploys more rigorous topological sampling for obtaining support values in addition to approximate likelihood-ratio tests.

Posterior probabilities

Posterior probabilities for bipartitions are obtained from Bayesian analyses, assuming that the parameter space has been sampled to a sufficient degree. The Metropolis [60] or more commonly the Metropolis-Hastings [61] Markov chain Monte Carlo (MCMC) algorithm must have run long enough to converge, and the chosen prior probabilities must be reasonable. Bipartition support values can then be obtained, for example, by building consensus trees from the posterior set of trees that have been sampled (using the bipartitions and posterior probabilities of the set of trees). The art of developing MCMC algorithms consists in designing proposal mechanisms that are able to efficiently (i.e. without spending many generations) escape local optima, avoiding early stopping of Bayesian analyses when chains reach apparent stationarity. While those problems can be alleviated by executing several chains, conducting multiple runs or using convergence analysis tools [62], there is still no guarantee that the tree search space has been sampled to a sufficient degree. In the worst case scenario, the topology proposals for one or more chains will almost always get rejected such that the same tree topology is sampled for slightly different model parameters (e.g. $\alpha$-shape parameter of the $\Gamma$ model of rate heterogeneity [31], GTR model parameters [30], branch lengths) that can then potentially lead to very high-posterior probabilities for incorrect tree bipartitions [63]. However, promising ideas to improve the proposal mechanism exist, such as the TBR-biased proposal in MrBayes 3.2 that allows the chain to more easily escape from situations of apparent stationarity (J. Huelsenbeck, personal communications). Clearly, Bayesian methods have advantages with respect to implementing more complex models such as mixture models [64, 65] (but see [66]) or heterotachy [67].

Figure 5: Outline of approximate likelihood-ratio tests for obtaining bipartition support values using NNI moves on the best-known NNI-optimal maximum likelihood tree.
that are more straightforward to integrate into a Bayesian (using a Gibbs sampler, see review in [68], or reversible jump MCMC algorithms [69]) than into a maximum likelihood framework. On the other hand, ML searches are more targeted in that they strive to find the absolute optimum in tree space and the search strategy may better fit the difficult NP-hard optimization problem. However, for ML there is also no guarantee that the search has recovered the global optimum. Finally, it has also been proposed to use bootstrapped posterior probabilities [70]; the authors also suggest that bootstrap proportions and plain posterior probabilities without bootstrapping should not be directly compared. See also [23, 33, 48, 70–73] for discussions and further references on the interpretation and comparison of bootstrap and posterior probability values.

PROGRAM VERIFICATION

Program verification represents a largely underestimated problem in disciplines, such as phylogenomics, that increasingly rely on scientific computing. Assuming that our methods and models are correct, in most cases, there is no guarantee that implementations are correct. While we are not aware of any catastrophic bugs in likelihood-based phylogeny programs, we will outline some of the potential pitfalls.

While theoretical tools for program verification such as for instance the Hoare calculus (also called Hoare logic or Hoare Rules) and software tools for program verification [74, 75] exist, they are rarely used in practice for academic software development (most popular tools for phylogenetic inference are freely available academic software) because of lack of time.

Likelihood-based programs face a substantial additional challenge, because they rely on floating point (also called machine numbers) arithmetics, whose correctness—if possible at all—is even more difficult to demonstrate [76], because simple mathematical rules for the real numbers such as \((a + b) + c = a + (b + c)\) do not hold for floating point arithmetics.

In fact, the problem is so difficult, that it required, for instance, an entire PhD thesis [77] to demonstrate that an implementation of elementary functions such as the exponential function, called millions of times in any phylogenetic likelihood implementation, returns correctly rounded results until the last bit under double-precision arithmetics.

Moreover, it is not guaranteed that the same code, for instance RAxML [11], will return exactly the same numerical results or even the same tree topology, if executed on different computer architectures, if executed sequentially or in parallel, or if compiled with different compilers. This behaviour results from compilers optimizing assembly code by assuming that the mathematical laws for real numbers hold for machine numbers. Additionally, many computer architectures allow for denormalized floating point values (i.e. floating point numbers that have more bits than the IEEE 754 standard), mainly to prevent numerical underflow.

The use of denormalized floating-point numbers also has serious implications on the assessment of supercomputer systems using floating-point intensive benchmark codes [78] because execution times can heavily depend upon the input data. This has been observed for the phylogenetic likelihood function during the development of the short-read phylogenetic placement algorithm in RAxML [79] where execution times for the likelihood function on data of the same size varied by 50% depending on the input data.

It would be possible to demonstrate the correctness of phylogenetic analysis programs that use parsimony or codes for reconstructing consensus trees because they operate on discrete entities, such as natural numbers, graphs and trees.

As a concrete example, for code verification-related erroneous conclusions in phylogenetics, a paper was published and later withdrawn, that assessed (using simulated data) the extent to which bootstrap-based bipartition support values were correlated with recovering true bipartitions. The paper stated that there was no correlation. However, a careful reader detected an error which pointed to a bug in the scripts that were used to analyze the bootstrap replicates and compute correlations. After fixing the bug, the conclusions of the paper were inverted and a strong correlation between bootstrap support values for bipartitions and their occurrence in the true simulated tree was observed. Unfortunately, the revised paper was not published to document the degree to which computational science and the hypotheses that are based on computational analyses rely on the correctness of software. Another example is, that the empirical base frequencies for C and G in the likelihood implementation of RAxML [11] were inverted until 2006 that is the frequency of C was used for G and vice versa. As a consequence,
RAXML returned topologically distinct trees when the difference between the empirical base frequencies of C and G was large.

To this end, extreme caution should be exercised with increasingly complex analysis pipelines that rely on a plethora of programs: sequence assembly, orthology assignment, MSA, tree building, post-analysis, biogeographical analyses and intermediate scripts to parse and adapt data formats. An example of the impact of program errors on scientific results is the retraction of five papers (published—among other journals—in Science and PNAS) on protein crystallography because of a bug in a data analysis program [80].

**CONCLUSIONS**

We have discussed issues related to the generally impossible task of verifying models, algorithms and software for phylogenetic inference and provided a brief overview on common techniques for obtaining support values on trees. There exist some promising empirical verification results using fast-evolving organisms [73, 81], which indicate that our reconstruction methods may be reasonable and that support values may be used for assessing whether a true bipartition has been recovered. However, this still represents anecdotal evidence and does not allow for extrapolation to larger time-scales (deeper phylogenies) or different types of organisms. Consider, for instance, the on-going debate about the ‘correct’ phylogeny of the metazoa [82–85]. As phylogenetics increasingly rely on computational methods, we believe that practitioners, should cease using Bioinformatics programs as black boxes and be aware of the important and potentially fatal effects software errors can have on the hypotheses they develop. While computational phylogenetics are coming off age, and widely used programs such as PhyloBayes, Beast, MrBayes, PHYML, RAXML, are becoming ever more complex from the software engineering point of view, more time should be dedicated to test and verify such codes (see also the discussion in [86] about the use of an incorrect Hastings ratio in several Bayesian programs).

**Key Points**

- Verification in phylogenetics is impossible, because the true evolutionary history of organisms is generally unknown.
- Given that the models are correct, there is no guarantee that the optimal tree, according to the model can be found, because

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